

Introduction to BLUPF90 software suite

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BLUPF90 TEAM – 03/2023



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BLUPF90 software suite

The State of Fortran

- Collection of software
 - Fortran \geq 90
 - Fortran = Formula Translation System
 - Fortran = Formula Translator
 - First compiler in 1957 by IBM

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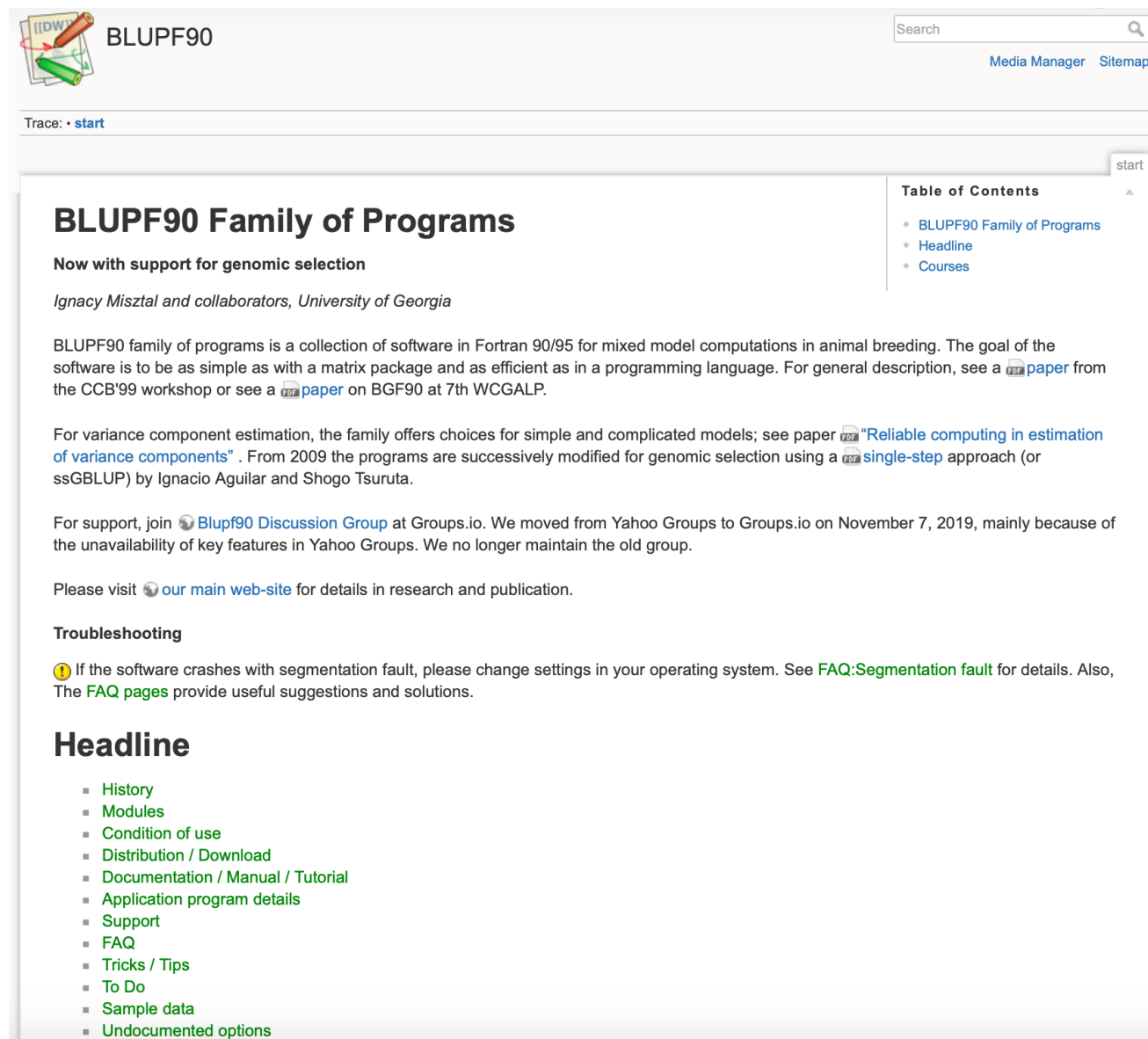
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Abstract—A community of developers has formed to modernize the Fortran ecosystem. In this article, we describe the high-level features of Fortran that continue to make it a good choice for scientists and engineers in the 21st century. Ongoing efforts include the development of a Fortran standard library and package manager, the fostering of a friendly and welcoming online community, improved compiler support, and language feature development. The lessons learned are common across contemporary programming languages and help reduce the learning curve and increase adoption of Fortran.

BLUPF90 software suite



The screenshot shows the BLUPF90 website homepage. At the top left is the BLUPF90 logo, which includes a stylized 'LDW' and a pencil. To the right of the logo is a search bar and links for 'Media Manager' and 'Sitemap'. Below the header, there is a 'Trace' section with a 'start' link. The main content area is titled 'BLUPF90 Family of Programs' and includes a sub-header 'Now with support for genomic selection' and the author information 'Ignacy Misztal and collaborators, University of Georgia'. The main text describes the software as a collection of Fortran 90/95 programs for mixed model computations in animal breeding. It also mentions a 'Table of Contents' sidebar with links to 'BLUPF90 Family of Programs', 'Headline', and 'Courses'. Below the main text, there are sections for 'Troubleshooting' and 'Headline' with a list of links: History, Modules, Condition of use, Distribution / Download, Documentation / Manual / Tutorial, Application program details, Support, FAQ, Tricks / Tips, To Do, Sample data, and Undocumented options.

- Collection of software
 - Fortran \geq 90
 - Computations in AB & G
- Since 1997/1998 by Ignacy Misztal
- Several developers + collaborators
- Simple, efficient, and comprehensive
 - Very general models

BLUPF90 software main developers



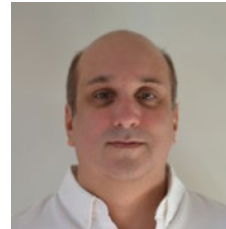
Ignacy
Misztal



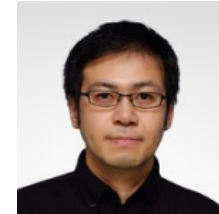
Shogo
Tsuruta



Andres
Legarra



Ignacio
Aguilar



Yutaka
Masuda



Matias
Bermann

- + Several contributors
- Research turns into code
- Which programs?

BLUPF90 software suite

blupf90

BLUP with explicit equations

remlf90

Expectation Maximization REML

airemlf90

Average Information REML

gibbsXf90

Bayesian Analyses – linear traits

thrgibbsXf90

Bayesian Analyses – categorical traits

nce.ads.uga.edu/wiki

Programs

Available for research (free)

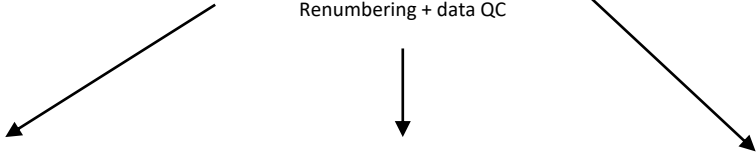
- **BLUPF90+** - a combined program of blupf90, remlf90, and airemlf90
- **GIBBSF90+** - a combined program of gibbs1f90, gibbs2f90, gibbs3f90, thrgibbs1f90, and thrgibbs3f90
- **POSTGIBBSF90** - statistics and graphics for post-Gibbs analysis (S. Tsuruta)
- **RENUMF90** - a renumbering program that also can check pedigrees and assign unknown parent groups; supports large data sets
- **PREGSF90** – genomic preprocessor that combines genomic and pedigree relationships (I. Aguilar)
- **POSTGSF90** – genomic postprocessor that extracts SNP solutions after genomic evaluations (single step, GBLUP) (I. Aguilar)
- **PREDICTF90** - a program to calculate adjusted y, y_hat, and residuals (I. Aguilar)
- **PREDF90** - a program to predict direct genomic value (DGV) for animals based on genotypes and SNP solution
- **QCF90** - a quality-control tool on genotypes and pedigree information (Y. Masuda)
- **INBUPGF90** - a program to calculate inbreeding coefficients with incomplete pedigree (I. Aguilar)
- **SEEKPARENTF90** - a program to verify paternity and parent discovery using SNP markers (I. Aguilar)

No longer updated (as of May 2022)

- **BLUPF90** - BLUP in memory
- **REMLF90** - accelerated EM REML
- **AIREMLF90** - Average Information REML with several options including EM-REML and heterogeneous residual variances (S. Tsuruta)
- **GIBBSF90** - simple block implementation of Gibbs sampling - no genomic
- **GIBBS1F90** - as above but faster for creating mixed model equations only once
- **GIBBS2F90** - as above but with joint sampling of correlated effects
- **GIBBS3F90** - as above with support for heterogeneous residual variances
- **THRGIBBSF90** - Gibbs sampling for any combination of categorical and linear traits (D. Lee) - no genomic
- **THRGIBBS1F90** - as above but simplified with several options (S. Tsuruta)
- **THRGIBBS3F90** - as above with heterogeneous residual variances for linear traits

renumf90

Renumbering + data QC



blupf90+

gibbsf90+

preGSf90

Processing of SNP data (QC + matrices)

qcf90 ✘

QC of large SNP data

postGSf90

Estimation of SNP effects and GWAS

predf90 ✘

Prediction of GEBV based on SNP effects

seekparentf90 ✘

Parentage verification (SNP and pedigree)

predictf90

Adjusted and predicted phenotypes + residuals

blup90iod2

blup90iod2OMP1

blup90iod3MPI1

cblup90iod2

cblup90iod2OMP1

accf90

accf90GS

blupf90

BLUP with explicit equations

remlf90

Expectation Maximization REML

airemlf90

Average Information REML

gibbsXf90

Bayesian Analyses – linear traits

thrgibbsXf90

Bayesian Analyses – categorical traits

postgibbsf90

Post-analyses of Gibbs samples

RENUMF90

**The renumbering software for the
BLUPF90 suite**

RENUMF90

- **Renumbers data and pedigree**
- **Creates a parameter file for BLUPF90 family**
 - **Parameter file can be modified by the users for new models**
- **Traces back pedigree for individuals in the data**
- **Performs comprehensive pedigree checks**
- **Provides data statistics**
- **Creates an Xref file for genotyped individuals**
- **Computes inbreeding by default in $v \geq 1.157$**



RENUMF90

- **Supports**

- virtually any dataset
- multiple traits
- different models (effects) per trait
- alphanumeric and numeric fields
- unknown parent groups
- covariates for random regression models

RENUMF90 – Input files

- **Data file and pedigree file as flat files**
 - Columns separated by at least one SPACE
 - No TABS !!!! (current version checks for it)
 - Input files cannot contain character #
 - Missing sire/dams must have code 0
 - code 00 is treated as a known animal

RENUMF90 – Output files

- **Creates files to be used by BLUPF90 family**
 - renf90.inb - file with inbreeding
 - renf90.tables - cross reference file with renumbered and original effects
 - renf90.fields - description of the effects in each field of renf90.dat

 - renf90.dat - renumbered data
 - renaddxx.ped - renumbered pedigree + statistics
 - renf90.par - new parameter file

RENUMF90 parameter file

MANDATORY

Keyword	possible value	description
DATAFILE	character	The name of data file to be processed
TRAITS	integer	Position for phenotype (trait) in the data file
FIELDS_PASSED TO OUTPUT	integer	Position for the columns in the original data that will be passed to the renumbered data without changes Keep empty if not needed
WEIGHT(S)	integer	The position(s) for weight in the data file Keep empty if not needed
RESIDUAL_VARIANCE	real value(s)	Residual (co)variance
EFFECT	(next slide ...)	Description of an effect Repeatable – 1 for each effect in the model

RENUMF90 parameter file

Effects

Keyword	Possible value	effect type	form
EFFECT	integer (column where the effect is)	cross	alpha
			numer
		cov	

Keyword (only for covariables)	Possible value	form
NESTED	integer (column where the effect is)	alpha
		numer

RENUMF90 parameter file

Model: $y = \text{farm} + \text{sex} + \beta \text{ age} + e$

```
DATAFILE
data1.txt
TRAITS
5
FIELDS_PASSED TO OUTPUT
2
WEIGHT(S)

RESIDUAL_VARIANCE
1.0
EFFECT      #1st effect - farm
2 cross alpha
EFFECT      #2nd effect - sex
3 cross numer
EFFECT      #3rd effect - age
4 cov
```

Fixed linear model

data1.txt

ID	farm	sex	age	phen
ID006	A	1	1.0	3.0
ID009	A	2	1.0	2.0
ID012	A	1	2.0	4.0
ID007	B	2	2.0	6.0
ID010	B	1	1.0	3.0
ID013	B	2	2.0	6.0
ID008	C	1	2.0	6.0
ID011	C	2	1.0	6.0
ID014	C	1	1.0	8.0
ID015	C	2	2.0	4.0

RENUMF90 parameter file

Random Effects

Keyword after EFFECT	possible value	description
RANDOM	diagonal	Non-correlated
	animal	Correlation structure among animals

Keyword	possible value	description
OPTIONAL	pe	Permanent environmental
	mat	Maternal
	mpe	Permanent environmental maternal (only if mat is used)

RENUMF90 parameter file

Random effects file section

Keyword after RANDOM (animal only)	possible value	description
FILE	character	Name of the pedigree file for animal models only

Keyword after FILE (for RANDOM animal only)	possible value	description
FILE_POS	integer	Specifies positions in the pedigree for ani sire dam alternate_dam yob Default: 1 2 3 0 0 <i>If maternal effect alternate_dam</i>

Keyword (for RANDOM animal only)	possible value	description
SNP_FILE	character	Optional: If genomic info is to be used Name of the SNP file Format: ID 011122211155152222

RENUMF90 parameter file

Pedigree options

Keyword (for RANDOM animal only)	possible value	description
PED_DEPTH	Integer	Optional Specifies the depth of pedigree search Default = 3 All pedigree = 0

RENUMF90 parameter file

Unknown Parent Group options

Keyword (for RANDOM animal only)	possible value	description
UPG_TYPE		Optional
	yob 1990 1992 ...	UPG assigned based on yob
	in_pedigrees	Missing parent receives -x x is the UPG number
	group_unisex	UPG based on the information in pedigree Ex. UPG by breed FILE_POS 1 2 3 0 0 4 #the 6th field indicates which column the UPG code is in the pedigree
group_sex	Separate UPG code for unknown sire and dam FILE_POS 1 2 3 0 0 4 5 #the 6 th and 7 th fields indicate which columns the UPG codes are in the pedigree	

RENUMF90 parameter file

Inbreeding option

Keyword (for RANDOM animal only)	possible value	description
INBREEDING	pedigree	Default in RENUMF90 \geq v1.157 Calculates inbreeding code and saves it in the renumbered pedigree file (Default in RENUMF90 \geq v1.157)
	File <name>	Reads inbreeding from an external file format: original_ID inbreeding (0 to 1)
	self \times	Calculates inbreeding with selfing \times is the column in the pedigree file with the number of selfing generations
	no-inbreeding	Turn inbreeding calculation off in RENUMF90 \geq v1.157

RENUMF90 parameter file

Random Regression options

Keyword	possible value	description
RANDOM_REGRESSION		Specifies that random regression should be applied to the random effects
	data	If covariables are in the data
	legendre	Generates legendre polynomials

Keyword	possible value	description
RR_POSITON	Integer	Specifies positions of covariables if RANDOM_REGRESSION type is data Specifies the order of the polynomial and the position of the covariable if RANDOM_REGRESSION type is legendre

RENUMF90 parameter file

(CO)VARIANCES for Random effects

Keyword	possible value	description
(CO)VARIANCES	real	(co)variance for the animal effect dimension should account for number of traits and random correlated effects

32.79	-7.22	-11.07
-7.22	258.06	87.66
-11.07	87.66	194.34

RENUMF90 parameter file

(CO)VARIANCES structure

- 3 trait (T1-T3) and 2 correlated effects (E1-E2)

		E1			E2		
		T1	T2	T3	T1	T2	T3
E1	T1						
	T2						
	T3						
E2	T1						
	T2						
	T3						

RENUMF90 parameter file

(CO)VARIANCES for Random effects

Keyword	possible value	description
(CO)VARIANCES	real	(co)variance for non-correlated random effects

Keyword	possible value	description
(CO)VARIANCES_PE	real	(co)variance for the PE effect if present

Keyword	possible value	description
(CO)VARIANCES_MPE	real	(co)variance for the MPE effect if present

RENUMF90 parameter file

Keyword	optional	possible values
COMBINE	optional	definition of new field as a combination of existing fields
DATAFILE	mandatory	name of raw data file
TRAITS	mandatory	positions of observations in the raw data file
FIELDS_PASSED	mandatory	positions of items in the raw data file to be passed to renf90.dat
WEIGHT(S)	mandatory	positions of weights in the raw data file
RESIDUAL_VARIANCE	mandatory	residual covariance matrix
EFFECT	mandatory	effect description
NESTED	optional	positions of nested covariates
RANDOM	optional	declaration of random effect
OPTIONAL	optional	declaration of MAT, PE, MPE
FILE	optional	name of raw pedigree file
FILE_POS	optional	positions of animal ID, sire ID, and dam ID
SNP_FILE	optional	name of SNP marker file
PED_DEPTH	optional	the maximum generation back from animals with phenotype and/or genotype
GEN_INT	optional	generation interval to set unknown parent groups (UPG)
REC_SEX	optional	check if records are found in specific sex
UPG_TYPE	optional	UPG specification
INBREEDING	optional	create pedigree file with inbreeding code
RANDOM_REGRESSION	optional	put covariates for random regressions
RR_POSITION	optional	positions of covariates for random regressions
(CO)VARIANCES	optional	covariance components
(CO)VARIANCES_PE	optional	covariance components for animal PE effects
(CO)VARIANCES_MPE	optional	covariance components for maternal PE effects
OPTION	optional	option parameters

RENUMF90 parameter file

- If the data and pedigree files have header

```
#Parameter file for renumf90
DATAFILE
data
SKIP_HEADER
1
TRAITS
3
FIELDS_PASSED TO OUTPUT
1 #Line_ID
WEIGHT(S)

RESIDUAL_VARIANCE
1.0
EFFECT
2 cross alpha
EFFECT
1 cross alpha
RANDOM
animal
FILE
ped
SKIP_HEADER
1
(CO)VARIANCES
1.0
OPTION sol se
```

RENUMF90 parameter file

Options passed to blupf90

- All lines that begin with the keyword `OPTION` are passed to parameter file `renf90.par`
 - Unless they are specific to `renumf90`
- This allows automation of process by using scripts
- For example:
 - `OPTION sol se`
 - `OPTION use_yams`

RENUMF90 parameter file

Hints

- Keyword EFFECT is repeated as many times as effects in the model
- If (CO)VARIANCES for any effect are missing, default matrix with 1.0 in diagonal and 0.1 on off-diagonal will be used

RENUMF90 parameter file

Model: $y = \text{farm} + \text{sex} + \beta \text{ age} + e$

```
DATAFILE
data1.txt
TRAITS
5
FIELDS_PASSED TO OUTPUT
2
WEIGHT(S)

RESIDUAL_VARIANCE
1.0
EFFECT      #1st effect - farm
2 cross alpha
RANDOM
diagonal
(CO)VARIANCES
0.5
EFFECT      #2nd effect - sex
3 cross numer
EFFECT      #3rd effect - age
4 cov
```

*What if we want to
consider farm as random
with variance = 0.5?*

data1.txt

ID	farm	sex	age	phen
ID006	A	1	1.0	3.0
ID009	A	2	1.0	2.0
ID012	A	1	2.0	4.0
ID007	B	2	2.0	6.0
ID010	B	1	1.0	3.0
ID013	B	2	2.0	6.0
ID008	C	1	2.0	6.0
ID011	C	2	1.0	6.0
ID014	C	1	1.0	8.0
ID015	C	2	2.0	4.0

RENUMF90 parameter file

Model: $y = \text{farm} + \text{sex} + \beta \text{ age} + \text{animal} + e$

```
DATAFILE
data1.txt
TRAITS
5
FIELDS_PASSED TO OUTPUT
2
WEIGHT(S)

RESIDUAL_VARIANCE
1.0
EFFECT      #1st effect - farm
2 cross alpha
EFFECT      #2nd effect - sex
3 cross numer
EFFECT      #3rd effect - age
4 cov
EFFECT      #4th effect - animal
1 cross alpha
RANDOM
animal
FILE
ped1.txt
FILE_POS
1 2 3 0 0
(CO)VARIANCES
0.2
```

*What if we want to consider
animal effect as random with
 $\sigma_u^2 = 0.2$?*

ped1.txt			data1.txt				
ID	Sire	Dam	ID	farm	sex	age	phen
ID006	ID001	ID003	ID006	A	1	1.0	3.0
ID009	ID001	ID004	ID009	A	2	1.0	2.0
ID012	ID001	ID005	ID012	A	1	2.0	4.0
ID007	ID001	ID003	ID007	B	2	2.0	6.0
ID010	ID001	ID004	ID010	B	1	1.0	3.0
ID013	ID002	ID005	ID013	B	2	2.0	6.0
ID008	ID002	ID003	ID008	C	1	2.0	6.0
ID011	ID002	ID004	ID011	C	2	1.0	6.0
ID014	ID002	ID005	ID014	C	1	1.0	8.0
ID015	ID002	ID003	ID015	C	2	2.0	4.0

RENUMF90 output files

Pedigree file: `renaddxx.ped`

Data file: `renf90.dat`

Parameter file: `renf90.par`

Inbreeding file: `renf90.inb`

Renumbering table: `renf90.table`

Fields table: `renf90.fields`

RENUMF90 output files

Pedigree file: `renaddxx.ped`

- Structure:

1. Animal ID (from 1)
2. Parent 1 ID or UPG number for parent 1
3. Parent 2 ID or UPG number for parent 2
4. 3 minus number of known parents
5. Known or estimated year of birth (0 if not provided)
6. Number of known parents
 if genotyped: 10+number of known parents
7. Number of records
8. Number of progeny as parent 1
9. Number of progeny as parent 2
10. Original animal ID

RENUMF90 output files

Pedigree file: `renaddxx.ped`

- As inbreeding is default:

Column 4:

$$\text{inb/upg code} = 4000 / [(1+m_s)(1-F_s) + (1+m_d)(1-F_d)]$$

m_s (m_d) is 0 if sire (dam) is known, and 1 otherwise

F_s (F_d) is the coefficient of inbreeding of sire (dam)

Ex: For an animal with both parents known and $F=0$

$$\text{inb/upg code} = 2000$$

RENUMF90 output files

Inbreeding file: `renf90.inb`

- `renf90.inb` will have:

origID	Inbreeding	newID
A71342462	0.059204	6927175
A17194772	0.032106	29
A13476873	0.002958	6550405
A1ZEP4813	0.000000	61
A14347077	0.019187	6550336
A64547711	0.026603	12
A71922414	0.000000	6942899
A17274771	0.019961	42
A53301967	0.000000	6550416
A4ZGF7566	0.000000	167
A3ZZS6645	0.000000	25
A07818367	0.000000	7117564
A17354770	0.050361	55
A53401908	0.000000	31
A13556872	0.063467	6550439
A14507075	0.071151	6550347

RENUMF90 output files

parameter file: renf90.par

```
# BLUPF90 parameter file created by RENUMF90
DATAFILE
  renf90.dat
NUMBER_OF_TRAITS
  1
NUMBER_OF_EFFECTS
  2
OBSERVATION(S)
  1
WEIGHT(S)

EFFECTS: POSITIONS_IN_DATAFILE NUMBER_OF_LEVELS TYPE_OF_EFFECT[EFFECT NESTED]
  2          2 cross
  3        12010 cross
RANDOM_RESIDUAL_VALUES
  0.60000
RANDOM_GROUP
  2
RANDOM_TYPE
  add_an_upginb
FILE
  renadd02.ped
(CO)VARIANCES
  0.40000
OPTION SNP_file genotypes.txt
OPTION map_file gen_map.txt
```

```
DATAFILE
  phenotypes.txt
TRAITS
  3
FIELDS_PASSED TO OUTPUT

WEIGHT(S)

RESIDUAL_VARIANCE
  0.60
EFFECT
  2 cross alpha #sex
EFFECT
  1 cross alpha
RANDOM
  animal
FILE
  pedigree.txt
FILE_POS
  1 2 3 0 0
SNP_FILE
  genotypes.txt
PED_DEPTH
  4
(CO)VARIANCES
  0.40
OPTION map_file gen_map.txt
```

renumf90 FAQ

1) renumf90 cannot find the data file

Check for typos

2) How to include quadratic covariable?

Column in data file

3) Error when trying to use covariable

2 cov numer

4) Fixed effects in renf90.dat are different from original

renf90.tables

5) I want to have original IDs in renf90.dat

FIELDS_PASSED TO OUTPUT

renumf90 FAQ

6) When and how to run renumf90?

a) Objective to compare models

Run renumf90 ONCE with the most complete model

Remove effects from renf90.par

b) Objective to compare non-genomic vs genomic model

Run renumf90 ONCE with SNP file

For non-genomic: Remove option for SNP file from renf90.par

c) Objective to mask phenotypes for some animals for validation

Run renumf90 ONCE with the complete data

Remove animals from renf90.dat

renumf90 quick trick

- `renumf90 --help`
- `renumf90 --show-template`

```
[dani@dodo2 day13]$ renumf90 --help
RENUMF90 version 1.158 with zlib

renumf90 parameter-file [--options ...]

--version          show version number
--show-template    show template parameter file
[dani@dodo2 day13]$ renumf90 --show-template
# parameter file for renumf90
DATAFILE

TRAITS

FIELDS_PASSED TO OUTPUT

WEIGHT(S)

RESIDUAL_VARIANCE

EFFECT

#RANDOM
#
#OPTIONAL
#
#FILE
#
#FILE_POS
#
#SNP_FILE
#
#PED_DEPTH
#
#UPG_TYPE
#
#INBREEDING
#
#FIXED_REGRESSION
#
#RANDOM_REGRESSION
#
#RR_POSITION
#
#(CO)VARIANCES
#
#(CO)VARIANCES_PE
#
#(CO)VARIANCES_MPE
#
#OPTION alpha_size 20
#OPTION max_string_readline 800
#OPTION max_field_readline 100
```

blupf90+

- blupf90: MME solver
- airemlf90: variance components using Average Information REML
- remlf90: variance components using Expectation Maximization REML

Mixed Model Equations Solver
Variance Components Estimation

$$\begin{bmatrix} \mathbf{X}'\mathbf{R}^{-1}\mathbf{X} & \mathbf{X}'\mathbf{R}^{-1}\mathbf{W} \\ \mathbf{W}'\mathbf{R}^{-1}\mathbf{X} & \mathbf{W}'\mathbf{R}^{-1}\mathbf{W} + \mathbf{A}^{-1} \otimes \mathbf{G}_0^{-1} \end{bmatrix} \begin{bmatrix} \hat{\boldsymbol{\beta}} \\ \hat{\mathbf{u}} \end{bmatrix} = \begin{bmatrix} \mathbf{X}'\mathbf{R}^{-1}\mathbf{y} \\ \mathbf{W}'\mathbf{R}^{-1}\mathbf{y} \end{bmatrix}$$

blupf90+



MME Solver

Default



VC Estimation

- AI-REML:

```
OPTION method VCE
```

- EM-REML:

```
OPTION method VCE
```

```
OPTION EM-REML xx
```

└─ # of EM rounds

xx > 0 : switch to aireml

xx < 0 : does not switch if convergence is reached

blupf90+

- Supports virtually any model used in AB&G:
 - animal model
 - models with maternal effect
 - MPE
 - PE
 - Random Regression
 - Social interaction
 - Multiple traits
 - up to 70 if no correlated effects
 - up to $\lfloor 70/\text{number of correlated effects} \rfloor$

blupf90+

- Computes generalized solutions by several methods:
 - Preconditioner Conjugate Gradient (PCG)
 - Default Iterative method (fast)
 - Successive over-relaxation (SOR)
 - an iterative method based on Gauss-Seidel
 - Direct solution using sparse Cholesky factorization
 - FSPAK or YAMS (greater memory requirements)
- Solutions change among methods, but estimable functions should be the same
- Prediction error variances can be obtained using sparse inverse (FSPAK or YAMS)

blupf90+ with PCG

Animal Breeding and Genetics Local Wiki

Iteration on data with preconditioned conjugate gradient (PCG)

Table of Contents

-

Algorithm

Preconditioned conjugate gradient (PCG) is an iterative method to solve the linear equations. This method is easily harmonized with the iteration of data technique. Intermediate status is kept in only 4 vectors and the one iteration will be done updating the vectors. BLUP90IOD2 is a program implementing the algorithms. Here we will introduce a basic idea needed to understand what the program does. See Stranden and Lidauer (2000) and Tsuruta et al. (2001) for detailed algorithm.

• Iteration on data with preconditioned conjugate gradient (PCG)
• Algorithm
• Programs
• Files and analysis
• Options

The mixed model equations can be written as

$$\mathbf{C}\mathbf{x} = \mathbf{b}$$

where \mathbf{C} is the left-hand side matrix, \mathbf{x} is the solution vector and \mathbf{b} is the right-hand side vector. If we have a matrix \mathbf{M} which is an approximation of \mathbf{C} , above equations are equivalent to

$$\mathbf{M}^{-1}\mathbf{C}\mathbf{x} = \mathbf{M}^{-1}\mathbf{b}.$$

This matrix \mathbf{M} is called preconditioner. If $\mathbf{M} = \mathbf{C}$, the equations are immediately solved. BLUPF90 uses $\mathbf{M} = \text{diag}(\mathbf{C})$ so its inverse is easily calculated.

The residual is expressed as

$$\mathbf{r} = \mathbf{b} - \mathbf{C}\mathbf{x}$$

and the algorithm tries to reduce with a statistics containing the residual. The **convergence** criterion is

$$\epsilon = \frac{\|\mathbf{b} - \mathbf{C}\mathbf{x}\|^2}{\|\mathbf{b}\|^2}$$

where $\|\cdot\|$ means the norm.

If $\mathbf{M}^{-1}\mathbf{C}$ has a better condition than \mathbf{C} , the convergence is reached is faster

Parameter file for blupf90+

```
# BLUPF90 parameter file created by RENUMF90
DATAFILE
  ../renf90.dat
NUMBER_OF_TRAITS      2
NUMBER_OF_EFFECTS    5
OBSERVATION(S)
  1 2
WEIGHT(S)

EFFECTS: POSITIONS_IN_DATAFILE NUMBER_OF_LEVELS TYPE_OF_EFFECT[EFFECT NESTED]
  3 4 40593 cross
  5 5 2 cross
  6 0 4 cross
  7 0 8 cross
  8 8 918111 cross
RANDOM_RESIDUAL VALUES
  2.5300 1.3425
  1.3425 29.714
RANDOM_GROUP
  5
RANDOM_TYPE
  add_an_upginb
FILE
  ../renadd05.ped
(CO)VARIANCES
  0.7600 2.2391
  2.2391 30.609
```

} Unlimited number of traits and effects

Parameter file for blupf90+

```
# BLUPF90 parameter file created by RENUMF90
```

```
DATAFILE
```

```
../renf90.dat
```

```
NUMBER_OF_TRAITS
```

```
2
```

```
NUMBER_OF_EFFECTS
```

```
5
```

```
OBSERVATION(S)
```

```
1 2
```

```
WEIGHT(S)
```

As many columns as the number of traits

Number of levels

```
EFFECTS: POSITIONS_IN_DATAFILE NUMBER_OF_LEVELS TYPE_OF_EFFECT[EFFECT NESTED]
```

```
3 4 40593 cross  
5 5 2 cross  
6 0 4 cross  
7 0 8 cross  
8 8 918111 cross
```

Type of effect

```
RANDOM_RESIDUAL VALUES
```

```
2.5300 1.3425
```

```
1.3425 29.714
```

```
RANDOM_GROUP
```

```
5
```

```
RANDOM_TYPE
```

```
add_an_upginb
```

```
FILE
```

```
../renadd05.ped
```

```
(CO) VARIANCES
```

```
0.7600 2.2391
```

```
2.2391 30.609
```

- As many rows as the NUMBER_OF_EFFECTS
- Model definition for each trait
- Different models per trait are supported
- If an effect is missing for one trait use 0

Parameter file for blupf90+

```
# BLUPF90 parameter file created by RENUMF90
DATAFILE
  ../renf90.dat
NUMBER_OF_TRAITS
  2
NUMBER_OF_EFFECTS
  5
OBSERVATION(S)
  1 2
WEIGHT(S)

EFFECTS: POSITIONS_IN_DATAFILE NUMBER_OF_LEVELS TYPE_OF_EFFECT[EFFECT NESTED]
  3 4 40593 cross
  5 5 2 cross
  6 0 4 cross
  7 0 8 cross
  8 8 918111 cross
RANDOM_RESIDUAL_VALUES
  2.5300 1.3425
  1.3425 29.714
RANDOM_GROUP
  5
RANDOM_TYPE
  add_an_upginb
FILE
  ../renadd05.ped
(CO)VARIANCES
  0.7600 2.2391
  2.2391 30.609
```

} Should be a square matrix with dimension equal to the number of traits

- Use zero (0.0) to indicate uncorrelated residual effects between traits
- e.g. For a 3-trait model
43.1 0.0 0.0
0.0 5.1 3.2
0.0 3.2 10.3

Parameter file for blupf90+

```
# BLUPF90 parameter file created by RENUMF90
DATAFILE
  ../renf90.dat
NUMBER_OF_TRAITS
  2
NUMBER_OF_EFFECTS
  5
OBSERVATION(S)
  1 2
WEIGHT(S)

EFFECTS: POSITIONS_IN_DATAFILE NUMBER_OF_LEVELS TYPE_OF_EFFECT[EFFECT NESTED]
  3 4 40593 cross
  5 5 2 cross
  6 0 4 cross
  7 0 8 cross
  8 8 918111 cross
RANDOM_RESIDUAL_VALUES
  2.5300 1.3425
  1.3425 29.714
RANDOM_GROUP
  5
RANDOM_TYPE
  add_an_upginb
FILE
  ../renadd05.ped
(CO) VARIANCES
  0.7600 2.2391
  2.2391 30.609
```

Definition of random effects

RANDOM_GROUP
RANDOM_TYPE
FILE
(CO) VARIANCES

Definition of random effects

- RANDOM_GROUP
 - Number of the effect(s) from list of effects
 - Correlated effects should be consecutive e.g. Maternal effects, Random Regression
- RANDOM_TYPE
 - diagonal, add_animal, add_sire, add_an_upg, add_an_upginb, add_an_self, user_file, user_file_i, or par_domin
- FILE
 - Pedigree file, parental dominance, or user file
- (CO)VARIANCES
 - Square matrix with dimension equal to the number_of_traits*number_of_correlated_effects

(CO)VARIANCES

- Assuming a 3 trait (T1-T3) and 2 correlated effects (E1-E2)

		E1			E2		
		T1	T2	T3	T1	T2	T3
E1	T1						
	T2						
	T3						
E2	T1						
	T2						
	T3						

RANDOM_TYPE

- *Diagonal*
 - for permanent environment effects
 - assumes no correlation between levels of the effect
- *add_sire*
 - To create a relationship matrix using sire and maternal grandsire
 - Pedigre file:
 - `individual number, sire number, maternal grandsire number`
- *add_animal*
 - To create a relationship matrix using sire and dam information
 - Pedigre file:
 - `animal number, sire number, dam number`

RANDOM_TYPE

- *add_an_upg*
 - As before but using rules for unknown parent group
 - Pedigree file:
 - animal number, sire number, dam number, parent code
 - missing sire/dam can be replaced by upg number, usually greater than maximum number of animals
 - Parent code = 3 – # of known parents
 - 1 both parents known
 - 2 one parent known
 - 3 both parents unknown
- *add_an_upginb*
 - As before but using rules for unknown parent group and inbreeding
 - Pedigree file:
 - animal number, sire number, dam number, inb/upg code
 - missing sire/dam can be replaced by upg number, usually greater than maximum number of animals
 - $\text{inb/upg code} = 4000 / [(1+m_s)(1-F_s) + (1+m_d)(1-F_d)]$
 - m_s (m_d) is 0 if sire (dam) is known and 1 otherwise
 - F_s (F_d) inbreeding coefficient of the sire (dam)

RANDOM_TYPE

- *Add_an_self*
 - To create a relationship matrix when there is selfing
 - Pedigre file:
 - individual number, parent 1 number, parent 2, number of selfing generations
- *user_file*
 - An inverted matrix is read from file
 - Matrix is stored only upper- or lower-triangular
 - Matrix file:
 - row, col, value
- *user_file_i*
 - As before but the matrix will be inverted by the program
- *par_domin*
 - A parental dominance file created by program RENDOM

OPTIONS for blupf90+

- Program behavior can be modified by adding extra options at the end of the par file
- `OPTION option_name x1 x2 ...`
- `option_name`: each program has its definition of options
- The number of optional parameters (`x1, x2, ...`) to control the behavior depends on the option

Options for blupf90+

Options

```
OPTION conv_crit 1e-12
```

Set convergence criteria (default 1e-12).

```
OPTION maxrounds 10000
```

Set maximum number of rounds (default 5000).

```
OPTION solv_method FSPAK
```

Selection solutions by FSPAK, SOR or PCG (default PCG).

```
OPTION r_factor 1.6
```

Set relaxation factor for SOR (default 1.4).

```
OPTION sol se
```

Store solutions and standard errors.

```
OPTION store_peg_pec 6
```

Store triangular matrices of standard errors and its covariances for correlated random effects such as direct-maternal effects and random-regression effects in "peg_pec_bf90".

Options for blupf90+

Missing data
Not pedigree!



```
OPTION missing -999
```

Specify missing observations (default 0) in integer.

```
OPTION residual
```

y-hat and residual will be included in "yhat_residual".

```
OPTION blksize 3
```

Set block size for preconditioner (default 1).

```
OPTION use_yams
```

Run the program with YAMS (modified FSPAK).

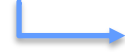
```
OPTION SNP_file snp
```

Specify the SNP file name to use genotype data.

New options for blupf90+

- Storing reliabilities based on PEV

OPTION store_accuracy X



Number of animal effect

$$Rel = 1 - \frac{PEV}{\sigma_u^2(1 + f)}$$

- Adjusts for f (inbreeding) from **A**, **G**, or **H**
 - Turn inbreeding adjustment off
 - OPTION correct_accuracy_by_inbreeding_direct 0
-
- Storing solutions with original ID if renumf90 was used to renumber the data
- OPTION origID
- Only *solutions.original* is created

Common parameter file for blupf90+

```
# BLUPF90 parameter file created by RENUMF90
DATAFILE
  renf90.dat
NUMBER_OF_TRAITS
  1
NUMBER_OF_EFFECTS
  2
OBSERVATION(S)
  1
WEIGHT(S)

EFFECTS: POSITIONS_IN_DATAFILE NUMBER_OF_LEVELS TYPE_OF_EFFECT[EFFECT NESTED]
  2          2 cross
  3      12010 cross
RANDOM_RESIDUAL_VALUES
  0.60000
RANDOM_GROUP
  2
RANDOM_TYPE
  add_an_upginb
FILE
  renadd02.ped
(CO)VARIANCES
  0.40000
OPTION SNP_file genotypes.txt
OPTION map_file gen_map.txt
```

blupf90+



VC Estimation

REML

- blupf90+ has 2 REML algorithms
 - EM-REML: expectation-maximization (EM) algorithm
 - AI-REML: average information (AI) algorithm
- REML = restricted/residual maximum likelihood
 - Patterson and Thompson (1971)
- Most used method for VCE in AB&G

EM-REML

- This method requires iterations:

$$\mathbf{y} = \mathbf{X}\boldsymbol{\beta} + \mathbf{Z}\mathbf{u} + \mathbf{e} \quad \begin{bmatrix} \mathbf{X}'\mathbf{X} & \mathbf{X}'\mathbf{Z} \\ \mathbf{Z}'\mathbf{X} & \mathbf{Z}'\mathbf{Z} + \mathbf{A}^{-1} \frac{\sigma_e^2}{\sigma_a^2} \end{bmatrix} \begin{bmatrix} \hat{\boldsymbol{\beta}} \\ \hat{\mathbf{u}} \end{bmatrix} = \begin{bmatrix} \mathbf{X}'\mathbf{y} \\ \mathbf{Z}'\mathbf{y} \end{bmatrix}$$

- 1) set initial variance components
- 2) compute $\hat{\boldsymbol{\beta}}$ and $\hat{\mathbf{u}}$ via mixed model equations
- 3) update variance components with the following equations

$$\hat{\sigma}_a^2 = \frac{\hat{\mathbf{u}}' \mathbf{A}^{-1} \hat{\mathbf{u}} + \text{tr}(\mathbf{A}^{-1} \mathbf{C}^{uu})}{N_a}$$

Inverse of LHS for animal effect

$$\hat{\sigma}_e^2 = \frac{\mathbf{y}'(\mathbf{y} - \mathbf{X}\hat{\boldsymbol{\beta}} - \mathbf{Z}\hat{\mathbf{u}})}{N - \text{rank}(\mathbf{X})}$$

animals (rank of A)

- 4) go to 1 or stop if the parameters do not change anymore

EM-REML

- Simpler equations
 - More complicated equations in multiple-trait models
- Easier to understand
- Very slow convergence (looks stable but may not converge)
- Computationally demanding, especially for C^{uu}

$$\begin{bmatrix} \hat{\beta} \\ \hat{u} \end{bmatrix} = \begin{bmatrix} \mathbf{X}'\mathbf{X} & \mathbf{X}'\mathbf{Z} \\ \mathbf{Z}'\mathbf{X} & \mathbf{Z}'\mathbf{Z} + \mathbf{A}^{-1} \frac{\sigma_e^2}{\sigma_a^2} \end{bmatrix}^{-1} \begin{bmatrix} \mathbf{X}'\mathbf{y} \\ \mathbf{Z}'\mathbf{y} \end{bmatrix}$$

AI-REML

Vector of variance components

$$\theta_{n+1} = \theta_n - \mathbf{H}^{-1}(\theta_n) \mathbf{d}(\theta_n)$$

Hessian Matrix

Gradient (score vector)

Average-information algorithm uses this matrix as Hessian,

P = Projection or hat matrix

$$\mathbf{H}(\theta) = \mathcal{I}_A(\theta) = \begin{bmatrix} -\frac{1}{2} \mathbf{y}' \mathbf{P} \mathbf{Z} \mathbf{A} \mathbf{Z}' \mathbf{P} \mathbf{Z} \mathbf{A} \mathbf{Z}' \mathbf{P} \mathbf{y} & -\frac{1}{2} \mathbf{y}' \mathbf{P} \mathbf{Z} \mathbf{A} \mathbf{Z}' \mathbf{P} \mathbf{P} \mathbf{y} \\ -\frac{1}{2} \mathbf{y}' \mathbf{P} \mathbf{P} \mathbf{Z} \mathbf{A} \mathbf{Z}' \mathbf{P} \mathbf{y} & -\frac{1}{2} \mathbf{y}' \mathbf{P} \mathbf{P} \mathbf{P} \mathbf{y} \end{bmatrix}$$

Gradient

expensive

$$-2\mathbf{d}(\theta) = \begin{bmatrix} \text{tr}(\mathbf{P} \mathbf{Z} \mathbf{A} \mathbf{Z}') - \mathbf{y}' \mathbf{P} \mathbf{Z} \mathbf{A} \mathbf{Z}' \mathbf{P} \mathbf{y} \\ \text{tr}(\mathbf{P}) - \mathbf{y}' \mathbf{P} \mathbf{P} \mathbf{y} \end{bmatrix} = \begin{bmatrix} \frac{N_a}{\sigma_a^2} - \frac{\text{tr}(\mathbf{A}^{-1} \mathbf{C}^{uu})}{(\sigma_a^2)^2} - \frac{\hat{\mathbf{u}}' \mathbf{A}^{-1} \hat{\mathbf{u}}}{(\sigma_a^2)^2} \\ \frac{N - \text{rank}(\mathbf{X})}{\sigma_e^2} - \frac{1}{\sigma_e^2} \left[N_a - \frac{\text{tr}(\mathbf{A}^{-1} \mathbf{C}^{uu})}{\sigma_a^2} \right] - \frac{\hat{\mathbf{e}}' \hat{\mathbf{e}}}{(\sigma_e^2)^2} \end{bmatrix}$$

AI-REML

- Computationally demanding
- Much faster than EM-REML
 - Fewer iterations
- Provides estimation of standard errors
- BUT
 - For complex models and poor starting values
 - Slow convergence
 - Parameter estimates out of the parameter space
 - In some cases, initial rounds with EM-REML may help

blupf90+



VC Estimation

- AI-REML:

OPTION method VCE

- EM-REML:

OPTION method VCE

OPTION EM-REML xx

└─ # of EM rounds

xx > 0 : switch to aireml

xx < 0 : does not switch if convergence is reached

Original options for
airemlf90 and remlf90
also work!

Options for blupf90+

```
OPTION se_covar_function <label> <function>
```

<label>

A name for a particular function (e.g., P1 for phenotypic variance of trait 1, H2_1 for heritability for trait 1, rg12 for genetic correlation between traits 1 and 2, ...).

<function>

A formula to calculate a function of (co)variances to estimate SD. All terms of the function should be written with no spaces.

Each term of the function corresponds to (co)variance elements and could include any random effects (G) and residual (R) (co)variances.

```
G_eff1_eff2_trt1_trt2
```

```
R_trt1_trt1
```

Examples:

```
OPTION se_covar_function P G_2_2_1_1+G_2_3_1_1+G_3_3_1_1+G_4_4_1_1+R_1_1
```

```
OPTION se_covar_function H2d G_2_2_1_1/(G_2_2_1_1+G_2_3_1_1+G_3_3_1_1+G_4_4_1_1+R_1_1)
```

```
OPTION se_covar_function rg12 G_2_2_1_2/(G_2_2_1_1*G_2_2_2_2)**0.5
```

SE for genetic parameters

```
#genetic, permanent, residual
ahat=c(
  0.11478,
  0.13552,
  0.25290,
  )
```

with AI matrix:

```
# inverse of AI matrix (Sampling Variance)
AI=matrix(c(
  0.16799E-05, -0.96486E-06, -0.82566E-08,
  -0.96486E-06, 0.96167E-06, -0.37113E-07,
  -0.82566E-08, -0.37113E-07, 0.10864E-06)
,ncol=3)
```

```
require(MASS)
b=mvrnorm(10000,ahat,AI)
> head(b)
      [,1]      [,2]      [,3]
[1,] 0.1146738 0.1357640 0.2529399
[2,] 0.1163889 0.1342926 0.2528479
[3,] 0.1166155 0.1344342 0.2525161
[4,] 0.1142085 0.1358928 0.2534974
[5,] 0.1136835 0.1361108 0.2530133
[6,] 0.1140485 0.1365707 0.2530573
```

heritability and its standard deviation:

```
h2=b[,1]/(b[,1]+b[,2]+b[,3])
sd(h2)
> 0.002318198
```

SE for genetic parameters

Houle and Meyer (2015):

Large-sample theory shows that maximum-likelihood estimates (including restricted maximum likelihood, REML) asymptotically have a multivariate normal distribution, with covariance matrix derived from the inverse of the information matrix, and mean equal to the estimated \mathbf{G} . This suggests that sampling estimates of \mathbf{G} from this distribution can be used to assess the variability of estimates of \mathbf{G} , and of functions of \mathbf{G} .

\mathbf{G} = additive genetic variance–covariance matrices

Does blupf90+ for VCE always converge?

- When the expected variance is very small, or the covariance matrix is close to non-positive definite, try the following starting values:
 - much smaller = 0.00001
 - much bigger = 1000
- If blupf90+ does not converge with AI-REML but converges with EM-REML with the same data set and the same model:
 - run EM-REML again but with a smaller starting value to check the estimate because it could be an artifact
 - use `OPTION EM-REML` inside blupf90+ as an initial point for AI-REML:
 - `OPTION EM-REML xx`

blupf90+ quick trick

- `blupf90+ --help`

```
[dani@dodo2 day13]$ blupf90+ --help
*****
*   BLUPF90+   *
*****

Computation of variance components, solutions, and s.e.
Default behavior avoids variance components estimation
For help about genomics, use blupf90+ --help-genomic

* OPTION SNP_file snp
  Specify the SNP file name to use genotype data.

* OPTION method VCE (default BLUP with blupf90 options)
  Run airemlf90 for variance component estimation (default running blupf90)

* OPTION conv_crit ld-12
  Convergence criterion (default ld-10)

* OPTION maxrounds 1000
  Maximum rounds (default 5000).
  When maxrounds=0, calculates BLUP without iterating REML and some statistics

* OPTION EM-REML 10
  Run EM-REML (REMLF90) for first 10 rounds (default 0).

* OPTION use_yams
  Run the program with YAMS (modified FSPAK). The computing time can be dramatically improved.

* OPTION tol ld-12
  Tolerance (or precision) (default ld-14) for positive definite matrix and g-inverse subroutines.
  Convergence may be much faster by changing this value.

* OPTION sol se
  Store solutions and those standard errors.

* OPTION origID
  Store solutions with original IDs.

* OPTION store_peg_pec 6
  Store triangular matrices of standard errors and its covariances for correlated random effects
  such as direct-maternal effects and random-regression effects in "peg_pec_bf90".

* OPTION residual
  y-hat and residuals will be included in "yhat_residual".

* OPTION missing -999
  Specify the missing value (default 0) in integer.

* OPTION constant_var 5 1 2 ...
  5: effect number
  1: first trait number
  2: second trait number
  implying the covariance between traits 1 and 2 for effect 5.

* More information:
  Application program details: http://nce.ads.uga.edu/wiki/doku.php?id=application\_programs
  BLUPF90 family manual: http://nce.ads.uga.edu/wiki/lib/exe/fetch.php?media=blupf90\_all7.60f
```

gibbsf90+

- `gibbs1f90`: stores single trait matrices once – fast for multi-trait models
- `gibbs2f90`: `gibbs1f90` with joint sampling of correlated effects – Maternal effects and RRM
- `gibbs3f90`: `gibbs2f90` with heterogeneous residual variance
- `thrgibbs1f90`: for linear-threshold models
- `thrgibbs3f90`: `thrgibbs1f90` with heterogeneous residual variance

Variance Components Estimation Mixed Model Equations Solver

$$\begin{bmatrix} \mathbf{X}'\mathbf{R}^{-1}\mathbf{X} & \mathbf{X}'\mathbf{R}^{-1}\mathbf{W} \\ \mathbf{W}'\mathbf{R}^{-1}\mathbf{X} & \mathbf{W}'\mathbf{R}^{-1}\mathbf{W} + \mathbf{A}^{-1} \otimes \mathbf{G}_0^{-1} \end{bmatrix} \begin{bmatrix} \hat{\boldsymbol{\beta}} \\ \hat{\mathbf{u}} \end{bmatrix} = \begin{bmatrix} \mathbf{X}'\mathbf{R}^{-1}\mathbf{y} \\ \mathbf{W}'\mathbf{R}^{-1}\mathbf{y} \end{bmatrix}$$

gibbsf90+



Linear

Default



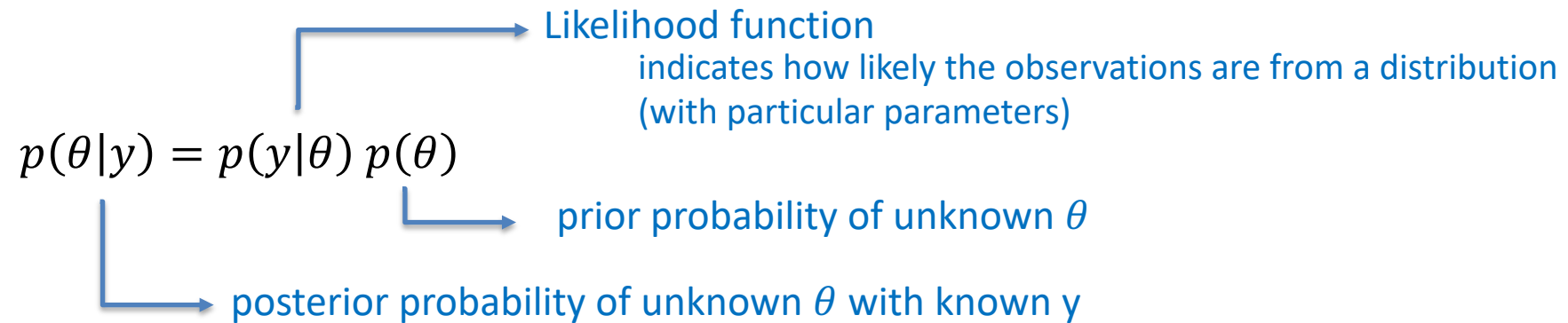
Threshold (-Linear)

```
OPTION cat 0 2 5
```

- Categories renumbered from **1**
- Missing records is only **0**

gibbsf90+

Bayes Theorem



- Basic idea of Gibbs Sampling:
- Numerical method to draw samples from a posterior distribution (not always explicitly available)
- Draw samples = generate random numbers following a distribution
- The results are random numbers (not theoretical formulas)
- The posterior distribution will be drawn based on the numerical values (like a histogram)

gibbsf90+

Ingredients for Gibbs sampling

- 1) Theoretical derivation: conditional posterior distribution for each unknown parameter
- 2) Software: a random number generator for a particular distribution

```
# Basic Gibbs sampling for mu (normal) and sigma2 (inverted chi-square)
y <- c(14,16,18)
N <- length(y)
n.samples <- 100
mu <- rep(0,n.samples)
sigma2 <- rep(0,n.samples)

# initial value
mu[1] <- 0
sigma2[1] <- 10

# sampling
for(i in 2:n.samples){
  mu[i] <- rnorm(1, mean=mean(y), sd=sqrt(sigma2[i-1]/N)) # using the most recent sigma2
  df <- N-2
  S <- sum((y-mu[i])^2)
  sigma2[i] <- rinvchisq(1, df=df, scale=S) # using the most recent mu
}
```

gibbsf90+

- Name of parameter file?

`gibbs1.par`

- Number of samples and length of burn-in?

`samples=10,000 to 100,000; burn-in=0`

- Give n to store every n-th sample?

`10`

- `gibbsf90+ parfile.par --samples i --burnin j --interval k`

gibbsf90+

- Procedure

- Run `gibbsf90+` to estimate variance components
- Run `postgibbsf90` to process the samples and check convergence
- Run `gibbsf90+` with new variance components to compute EBV (2k to 10k samples)

```
OPTION fixed_var mean X
```



Number of the
animal effect

gibbsf90+

```
OPTION cat 0 0 2 5
```

"0" indicate that the first and second traits are linear. "2" and "5" indicate that the third and fourth traits are categorical with 2 (binary) and 5 categories.

```
OPTION fixed_var all
```

Store all samples for solutions in "all_solutions" and posterior means and SD for all effects in "final_solutions", assuming that (co)variances in the parameter file are known.

```
OPTION fixed_var all 1 2 3
```

Store all samples for solutions in "all_solutions" and posterior means and SD for 1, 2, and 3 effects in "final_solutions", assuming that (co)variances in the parameter file are known.

```
OPTION fixed_var mean
```

Only posterior means and SD for solutions are calculated for all effects in "final_solutions", assuming that (co)variances in the parameter file are known.

```
OPTION fixed_var mean 1 2 3
```

Only posterior means and SD for solutions are calculated for effects 1, 2, and 3 in "final_solutions", assuming that (co)variances in the parameter file are known.

gibbsf90+

```
OPTION save_halfway_samples n
```

This option can help the 'cold start' (to continue the sampling when the program accidentally stops before completing the run). An integer value n is needed. In every n rounds, the program saves intermediate samples to 2 files (`last_solutions` and `binary_final_solutions`). The program can restart the sampling from the last round where the intermediate files were saved. The program also writes a log file `save_halfway_samples.txt` with useful information for the next run.

To restart, add `OPTION cont 1` to your parameter file and run `gibbsf90+` again. Input 3 numbers (samples, burn-in, and interval) according to `save_halfway_samples.txt`. Gibbsf90+ can take care of all restarting process by itself, so no other tools are needed.

Tips

- Small n will make the program slow because of frequent file writing. The n should be a multiple of the interval (the 3rd number you will input in the beginning of the program).
- If the program stops during burn-in, the restart will fail because `gibbs_samples` is not created. Recommendation is burn-in=0 (but it doesn't provide posterior mean and SD for solutions).
- The cold start may add tiny numerical errors to the samples. Samples from the cold start wouldn't be identical to samples from a non-stop analysis.
- If, unfortunately, the program is killed during its saving the intermediate samples, the cold start will fail. To avoid this, you can manually make a backup for `gibbs_samples`, `fort.99`, `last_solutions`, and `binary_final_solutions` at some point and write them back if needed.

```
OPTION se_covar_function <label> <function>
```

gibbsf90+

```
OPTION hetres_int col nlev
```

```
OPTION hetres_int 5 10
```

The position "5" to identify the interval in the data file and the number of intervals "10" for heterogeneous residual variances.

gibbsf90+

Data (datasire)

```
1 - HYS
2 - sire
3 - y1
4 - heterogeneous clas
5 - y2
```

cat datasire

```
6 13 317.55 1 644.26
3 10 280.44 1 563.05
.....
37 1 270.52 5 543.63
53 10 286.43 5 579.84
```

Parameter file (ex5)

```
DATAFILE
datasire
NUMBER_OF_TRAITS
NUMBER_OF_EFFECTS
OBSERVATION(S)
WEIGHT(S)
EFFECTS: POSITIONS_IN_DATAFILE
1 1 100 cross
2 2 50 cross
RANDOM_RESIDUAL VALUES
500 100
100 1000
RANDOM_GROUP
RANDOM_TYPE
diagonal
FILE
(CO)VARIANCES
75 10
10 150
OPTION hetres_int 4 5
```

```
round 98
209. 416.
416. 828.
Residual variance, interval 1
df_r 1997 ee/n 99.4738134864675
101. 202.
202. 412.
Residual variance, interval 2
df_r 1997 ee/n 146.518188769043
148. 296.
296. 602.
Residual variance, interval 3
df_r 1997 ee/n 198.183671561078
198. 397.
397. 806.
Residual variance, interval 4
df_r 1997 ee/n 232.307903786663
228. 455.
455. 917.
Residual variance, interval 5
df_r 1997 ee/n 301.189371418363
311. 622.
622. 0.126E+04
```

gibbsf90+ quick trick

- `gibbsf90+ --help`

```
[dani@dodo2 day13]$ gibbsf90+ --help
*****
*   GIBBSF90+   *
*****

Gibbs sampler for mixed threshold-linear models involving multiple categorical
and linear variables.
Thresholds and variances can be estimated or assumed.
For help about genomics, use gibbsf90+ --help-genomic

* OPTION SNP_file snp
  Specify the SNP file name to use genotype data.

* OPTION cat 0 0 2 5
  "0" indicate that the first and second traits are linear.
  "2" and "5" indicate that the third and fourth traits are categorical with 2 (binary) and 5 categories.

* OPTION fixed_var all
  Store all samples for solutions in all_solutions and posterior means and SD for all effects in final_solutions
  This assumes that (co)variances in the parameter file are known.

* OPTION fixed_var all 1 2 3
  Store all samples for solutions in all_solutions and posterior means and SD for 1, 2, and 3 effects in final_solutions
  This assumes that (co)variances in the parameter file are known.

* OPTION fixed_var mean
  Only posterior means and SD for solutions are calculated for all effects in final_solutions
  This assumes that (co)variances in the parameter file are known.

* OPTION fixed_var mean 1 2 3
  Only posterior means and SD for solutions are calculated for effects 1, 2, and 3 in final_solutions
  This assumes that (co)variances in the parameter file are known.
```


gibbsf90+ quick trick II

- Optimizing gibbsf90+ when using genomic data

Run renumf90 with the following option:

```
OPTION animal_order genotypes
```

Run gibbsf90+ with the following option:

```
OPTION separate_dense
```

postgibbsf90

- Basic idea of post-Gibbs analysis:
- Summarize and visualize the samples drawn by gibbsf90+
- Confirm if the chain converged
- Find the most probable value = posterior mode as a “point estimate”
- Find the reliability of the estimates = the highest posterior density as a “confidence interval”

postgibbsf90

- Name of parameter file?
gibbs1.par
- Burn-in?
0
- Give n to store every n-th sample? (1 means read all samples)
10
- input files
gibbs_samples, fort.99
- output files
 - "postgibbs_samples"
all Gibbs samples for additional post analyses
 - "postmean"
posterior means
 - "postsd"
posterior standard deviations
 - "postout"

postgibbsf90

at least > 10 is recommended
> 30 may be better

					*****	Monte	Carlo	Error by	Time Series	*****			
Pos.	eff1	eff2	trt1	trt2	MCE	Mean	HPD	Interval (95%)	Effective sample size	Median	Mode	Independent chain size	
1	4	4	1	1	1.362E-02	0.9889	0.7788	1.215	70.4	0.9844	0.9861	18	
2	4	4	1	2	1.288E-02	1.006	0.777	1.219	84.1	1.006	0.952	18	
3	4	4	2	2	1.847E-02	1.66	1.347	1.987	80.3	1.652	1.579	25	
4	0	0	1	1	9.530E-03	24.47	24.07	24.84	425.6	24.47	24.53	2	
5	0	0	1	2	8.253E-03	11.84	11.54	12.18	395.8	11.83	11.82	2	
6	0	0	2	2	1.233E-02	30.1	29.65	30.58	387.8	30.09	29.97	5	

***** p_i Lower and upper bounds of Mean ± 1.96PSD io ratio between first half and second half of the samples ; should be < 1.0

Pos.	eff1	eff2	trt1	trt2	PSD	Mean	PSD Interval (95%)	Geweke diagnostic	Autocorrelations lag: 1	10	50	Independent # batches
1	4	4	1	1	0.1144	0.9889	0.7648 1.213	-0.02	0.853	0.188	0.049	50
2	4	4	1	2	0.1182	1.006	0.7742 1.237	-0.11	0.828	0.111	-0.066	50
3	4	4	2	2	0.1656	1.66	1.335 1.984	0.06	0.828	0.108	-0.021	36
4	0	0	1	1	0.1967	24.47	24.09 24.86	-0.01	0.034	0.029	-0.062	450
5	0	0	1	2	0.1643	11.84	11.51 12.16	0.03	0.032	-0.006	-0.016	450
6	0	0	2	2	0.2429	30.1	29.62 30.57	-0.02	0.07	-0.014	0.037	180

postgibbsf90

```
Choose a graph for samples (= 1) or histogram (= 2); or exit (= 0)
```

```
1
```

```
positions
```

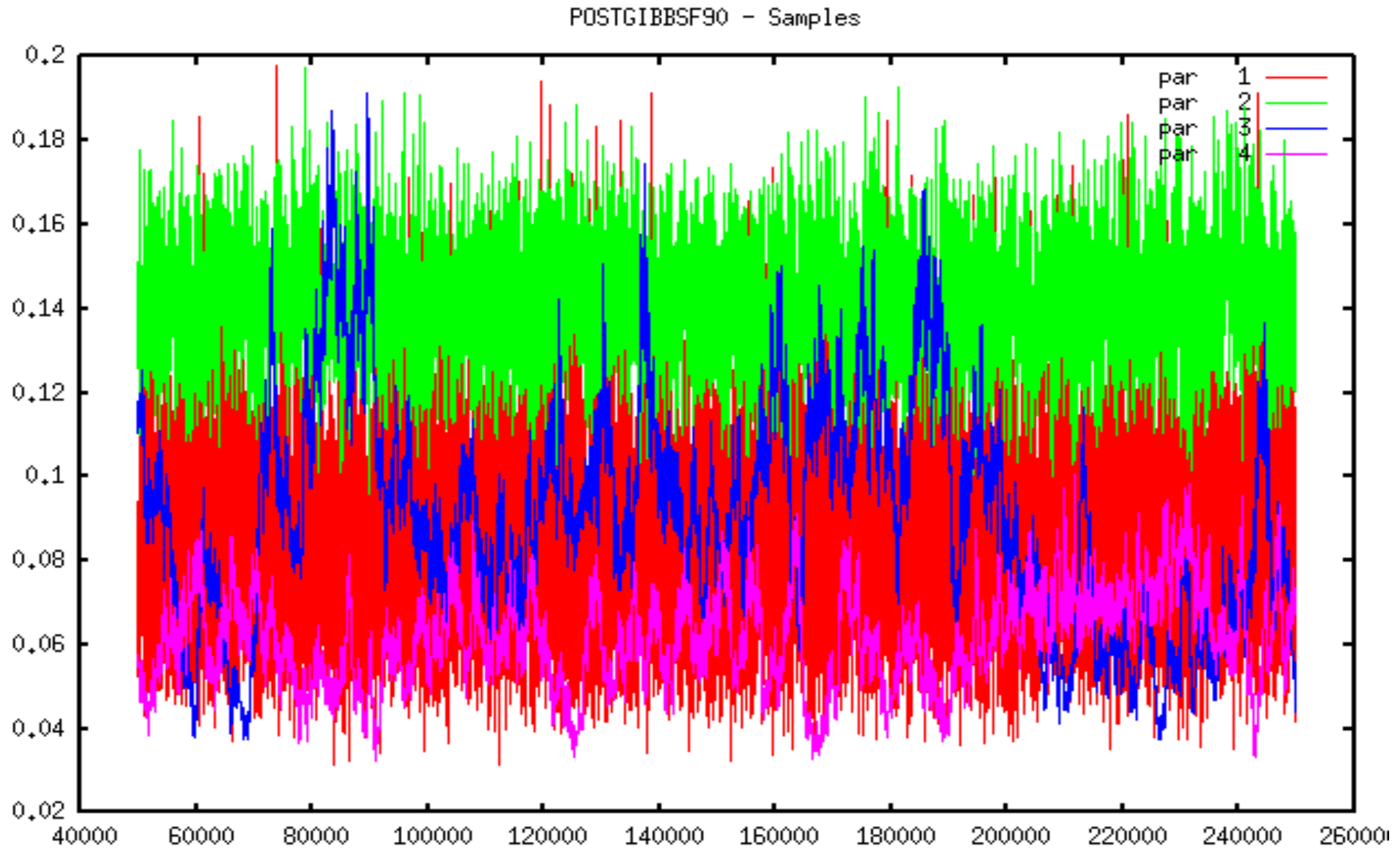
```
1 2 3 # choose from the position numbers 1 through 6
```

```
If the graph is stable (not increasing or decreasing), the convergence is met.  
All samples before that point should be discarded as burn-in.
```

```
print = 1; other graphs = 2; or stop = 0
```

```
2
```

postgibbsf90



postgibbsf90

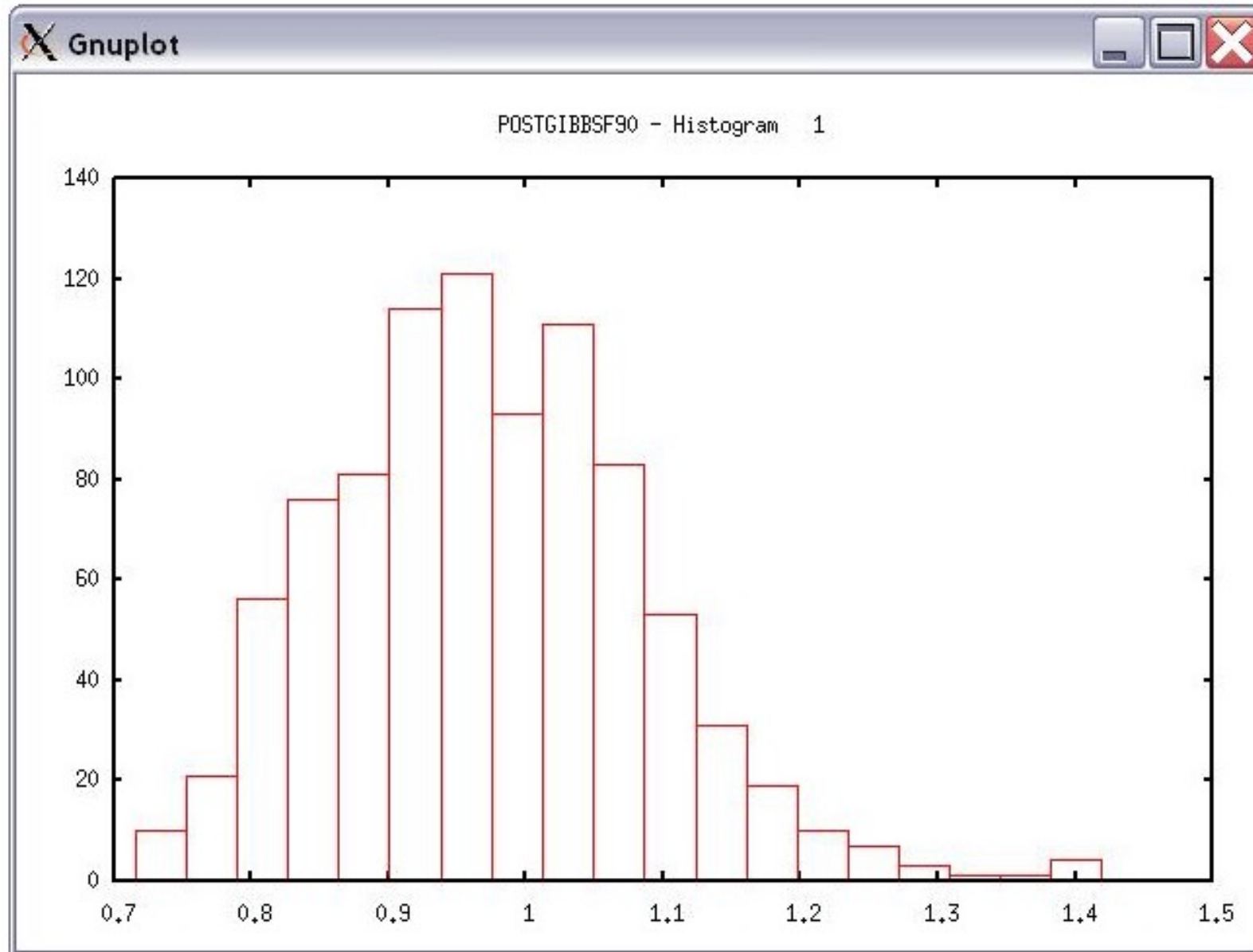
```
Choose a graph for samples (= 1) or histogram (= 2); or exit (= 0)
```

```
2
```

```
Type position and # bins
```

```
1 20
```

postgibbsf90



Common problems for BLUPF90 family

- Wrong position or formats for observation and effects
- Misspelling of Keywords
 - Program may stop
- (Co)variance matrices not symmetric, not positive definite
 - Program may not stop
- Large numbers (e.g., 305-day milk yield 10,000 kg)
 - Scale down i.e., $10,000 / 1,000 = 10$

General output from BLUPF90 family

- Output printed on the screen is not saved to any file!
- Should use redirection or pipes to store output

renumf90

```
renumf90 renum.par | tee renum.log
```

blupf90+

```
blupf90+ renumf90.par | tee blupf90.log
```

gibbsf90+

```
gibbsf90+ exmr99s1 --samples 1000 --burnin 0 --interval 1 | tee gibbsf90.log
```

Run in background + Save output

```
$vi gibbs.sh
#type the following commands inside gibbs.sh
    gibbsf90+ <<AA > gibbs.log
    renf90.par
    1000 0
    10
    AA
#save and exit
$bash gibbs.sh & #can replace bash with sh
```

```
$vi bp.sh
#type the following commands inside bp.sh
    blupf90+ <<AA > blup.log
    renf90.par
    AA
#save and exit
$bash bp.sh & #can replace bash by sh
```

Add Windows PATH Environment Variable

- Settings
- Control panel
- Search for advanced
- View advanced system settings
- Environmental variables
- System variables -> select **Path** -> [Edit...]
- ;C:\blupf90 (this is the path to the directory where programs are)
- Open a new terminal